



Fractional CO₂ laser for genitourinary syndrome of menopause in breast cancer survivors: clinical, immunological, and microbiological aspects

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Abstract

The composition of vaginal microbiome in menopause and cancer survivor women changes dramatically leading to genitourinary syndrome of menopause (GSM) in up to 7% of patients. Recent reports suggest that laser therapy may be valuable as a not hormonal therapeutic modality. The aim of the present study was to evaluate the effects of fractional CO₂ laser treatment on the vaginal secretory pathway of a large panel of immune mediators, usually implicated in tissue remodeling and inflammation, and on microbiome composition in postmenopausal breast cancer survivors. The Ion Torrent PGM platform and the Luminex BioPlex platform were used for microbiome and immune factor analysis. The significant reduction of clinical symptoms and the non-significant changes in vaginal microbiome support the efficacy and safety of laser treatment. Moreover, the high remodeling status in vaginal epithelium is demonstrated by the significant changes in inflammatory and modulatory cytokine patterns. Laser therapy can be used for the treatment of GSM symptoms and does not show any adverse effects. However, further studies will be needed to clarify its long-term efficacy and other effects.

Keywords Fractional CO₂ laser · Cytokines/chemokines · Microbiome · Menopause · Breast cancer

Abbreviations

| | |
|------|-------------------------------------|
| VVA | Vulvovaginal atrophy |
| GSM | Genitourinary syndrome of menopause |
| VRS | Verbal Rating Scale |
| BV | Bacterial vaginosis |
| VHI | Vaginal Health Index |
| FSFI | Female Sexual Function Index |

FSDSr Female Sexual Distress Scale revised 2005

Introduction

In menopause, the lack of estrogen is associated with diminished collagen, elastin, hyaluronic acid content, thinned

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epithelium, impaired smooth muscle proliferation, denser connective tissue arrangement, loss of vascularity, and impaired prolubricative and proelastic functions. The anatomical and functional defects predispose to irritation and sexual trauma and may lead to changes in sexual and urinary organs [1]. The classical symptoms are dyspareunia and dysuria, sometimes accompanied by recurrent urinary tract infections and other symptoms, eventually characterizing the syndrome called vulvovaginal atrophy (VVA) or, according to the most recent classification, genitourinary syndrome of menopause (GSM) [2]. All of these symptoms significantly affect the quality of life of menopausal women [3]. In breast cancer survivors, atrophic vaginitis/vaginal dryness can affect up to 70% of the postmenopausal patients when compared to postmenopausal women without breast cancer [4, 5]. The use of hormonal therapies improves menopausal syndrome, sexual health, and quality of life [1], although in postmenopausal women with a history of estrogen-dependent neoplasia, vaginal estrogen therapy is controversial and the most recent guidelines recommend co-management with the woman's oncologist [6]. In addition, breast cancer survivors, after surgery, usually receive further chemo-, hormonal, and radiotherapy that can aggravate the vaginal symptoms. In these patients, moisturizing treatments or lubricants are safe and indicated, even if they carry low compliance and they are less effective than hormonal therapies [7]. In the last few years, it has been suggested that laser therapy may be valuable as a non-hormonal therapeutic modality in the management of GSM [8]. There are approximately 15 laser companies in the market, the majority providing products based on the CO₂ and erbium:yttrium-aluminum-garnet (Er:YAG) lasers, and the interest in the use of these lasers as a non-hormonal option for the treatment of GSM has [9, 10].

Zerbinati et al. [11] have shown that the use of the micro-ablative carbon dioxide laser on the vaginal epithelial resurfacing activates heat shock proteins that in turn activate growth factors, stimulating the increase of acidic mucopolysaccharides in the ground matrix and the increase of glycogen content in the vaginal epithelial cells, with an increasing thickness of vaginal epithelium. Further studies have shown that the restoring of correct composition of the extracellular matrix produces a correct permeability of the connective favoring the passage of the different nutrients from the capillaries to the tissues. Particularly, the circulating estrogens produced in this phase in the adrenal gland and adipose tissue reach vaginal epithelial cells, which, since do not lose estrogen receptors in menopause, are stimulated to proliferate also with low estrogenic levels [12, 13].

Salvatore et al. [14] recently have published a pilot study on the treatment of VVA in postmenopausal women using fractional CO₂ laser where they observed morphological changes of collagen fibers in the vaginal epithelial tissue after treatment; these effects were associated with improvement of

VVA symptoms and patient quality of life. In particular, preliminary results of laser therapy efficacy have been reported on specific symptoms of VVA, such as sexual function [14] and vulvovaginal atrophy [15], vaginal flora [16], and GSM as a whole [17], and improvements were observed on women with idiopathic vestibulodynia or caused by GSM [18].

Accordingly with previous studies, our research team has shown the efficacy of the thermo-ablative fractional CO₂ laser in relieving dyspareunia symptoms in breast cancer survivor patients [19].

In the last few years, a growing body of evidence has suggested that vaginal health depends on a balance of several factors where the host local immunity and microbiome play a key role. Particularly, the composition of vaginal microbiome changes in menopause [20–22] and it is additionally modified by cancer survivor status could be some of the factors that cause GSM. A number of animal and human studies have shown that the levels of cytokines in cervicovaginal lavage (CVL) positively correlate with histopathology and clinical signs of vaginal inflammation, and that estrogens influence the activity of cytokines in the reproductive tract [23–26]. In the light of all these evidences, the aim of the present study was to evaluate the effects of laser treatment on vaginal immune mediators and microbiome of breast cancer women. Specifically, a secretory pathway of a large panel of immune mediators, usually implicated in tissue remodeling and inflammation, and microbiome composition were analyzed in postmenopausal cancer survivor women after vaginal mucosa treatment with fractional CO₂ laser.

Materials and methods

Patients

This is a clinical prospective study in consecutive postmenopausal patients who were referred to our clinic from December 2015 to February 2016 for one or more symptoms and signs related to GSM.

All women were postmenopausal and breast cancer survivors, and laser treatment was chosen as preferred therapy for vaginal symptoms as opposed to hormonal therapy, given the sensitivity of this population to estrogen levels. All patients underwent vaginal treatment with fractional CO₂ laser.

The inclusion criteria consisted of menopausal status, diagnosis of vaginal atrophy, patients with previous breast cancer, and PAP tests negative performed within 1 year of selection visit.

The diagnosis of vaginal atrophy was placed if there were the following signs and symptoms:

- “Vaginal dryness” as a symptom reported by the woman

- At least a sign of the following documented by the physician: flattening of the vaginal folds, dryness of the vaginal mucosa, pallor of the vaginal mucosa, fragility of the mucosa, and petechiae
- Vaginal pH > 5

Patients with active genital infections (bacterial vaginosis, *candida* vaginitis, and aerobic vaginitis), patients with clinically significant systemic diseases that may interfere with the study, patients with previous cancer of the vagina, cervix, or endometrium, patients previously treated with radiotherapy local, and/or the use of therapies vaginal moisturizers or lubricants in place were excluded from the study.

All the patients had completed the cancer treatment protocol and final evaluation within the study time.

All patients had discontinued therapy local vaginal moisturizing and/or lubricating at least 30 days, and the reasons for discontinuation of treatment were as follows: “Cream and/or vaginal suppositories tend to get out of the vagina” for seven patients; “therapies do not seem effective” for six patients; one had discontinued treatment for “local irritation”; one patient said that the vaginal applicator “is not easy to use,” and five patients were offered laser treatment as first choice.

The study was approved by the Ethics Committee of Florence. All patients who were recruited for the study signed an informed consent form. The study was conducted in accordance with the Declaration of Helsinki. This study was supported by the Italian Ministry of Health, grant number RC 26/13.

Laser treatment

Each patient was treated with the fractional CO₂ laser system (SmartXide2 V2LR, Monalisa Touch®, DEKA, Florence, Italy) using the vaginal probe at the Service of Laser Surgery and Colposcopy Daimi, AOU Careggi, Florence. All laser application are performed in an outpatient setting, and they did not require local anesthesia or other preparation. During the first and second treatment, the laser energy is set at 30-W power and transmitted through an intravaginal probe with a dwell time of 1000 μs, a dot spacing of 1000 μm, and a smart stack parameter of 1, following a protocol of application used in previous authors’ studies [27]. Laser energy is released by two single shots oriented at 45° one from the other so to treat the entire circular vaginal surface covered by the probe which is dotted of two opposite focusing points, without touching the uterine cervix [19]. These parameters were selected on the basis on ex vivo studies conducted on vaginal wall specimen, which demonstrated connective tissue remodeling [28]. The total exposure time for patient was approximately 3–5 min.

Clinical evaluation

The clinical evaluation of efficacy was performed individually for each of the following parameters which were carried during the screening visit (T0) and after 30 days from the second laser treatment (T1):

- Positioning of the speculum and observation of the vagina; signs of VVA were evaluated (flattening of the vaginal folds, dryness of the vaginal mucosa, pallor of the vaginal mucosa, fragility of the mucosa, petechiae) using a Verbal Rating Scale (VRS). Each parameter was graded from 0 (no symptom) to 3 (serious symptom)
- Vaginal swab for culture of bacteria and *Candida* spp. and Gram staining (Nugent score) for diagnosis of bacterial vaginosis (BV)
- Vaginal Health Index (VHI), which consisted of the following five parameters: elasticity, fluid volume, pH, epithelial integrity, and moisture. Each parameter was graded from 1 (worst condition) to 5 (best condition)
- Vaginal swab for the study of vaginal immunity and microbiome: the swab was scraped on the vaginal wall side (left and right) and then stored in a test tube with inside physiologic solution at – 20 °C
- Intensity of VVA symptoms was evaluated (vaginal itching, vaginal burning, vaginal dryness, dyspareunia, and dysuria) using a VRS. Each parameter was graded from 0 (no symptom) to 3 (serious symptom)
- Survey on the sexual function (the Female Sexual Function Index—FSFI) and survey on the sexual distress (the Female Sexual Distress Scale revised 2005—FSDSr).

At T1, treatment satisfaction was evaluated using a 5-point Likert scale (very satisfied, satisfied, uncertain, dissatisfied, and very dissatisfied).

Eventual adverse events which occurred or were referred by patients (during, immediately after treatment sessions, and until the end of study) were recorded.

Statistical analysis

Statistical analysis was performed using SPSS version 22.0 (Chicago, IL). The comparison between the variables mentioned above, before and after treatment with the laser fractionated CO₂, was performed by non-parametric paired Wilcoxon test or not paired Mann-Whitney test, as indicated. As a test of normality, the Shapiro-Wilk test was used. Shannon diversity (*H*) and equitability index were calculated and compared between the microbiomes before and after laser treatment. Statistical significance was set at $p < 0.05$. All the variables were not normally distributed.

Microbiome analysis

DNA extraction from the aliquots of 500 μ l was carried out using the NucliSENS® easyMAG® system (BioMérieux, Gorman, NC, USA). All DNA samples were stored at -20°C prior to Ion Torrent sequencing. A real-time quantitative EvaGreen® dye (Fisher Molecular Biology, Waltham, USA) PCR was performed with the degenerated primer 27FYM (5'-AGR GTT YGA TYM TGG CTC AG-3') and with the U534R primer (primers target the V1–V3 region, spanning 500 bp). A nested PCR was performed with the primers B338F_P1-adaptor (B338F 5'-ACTCCTACGGGAGGCAGC-3') and U534R_A_barcode (U534R 5'-ATTACCGCGGCTGCTGG-3') to prepare a 200 base template for the final V3 region sequencing, in association with the Ion Xpress Barcode Adapter. The PCR reactions were performed using the Kapa 2G HiFi Hotstart ready mix 2 \times (Kapa Biosystems, MA, USA) and BSA 400 ng/ μ l, under the following conditions: 5 min at 95°C , 30 s at 95°C , 30 s at $59^{\circ}/57^{\circ}\text{C}$, 45 s at 72°C , and a final elongation step at 72°C for 10 min. The size of the amplicon (260 bp) was checked on a 5% acrylamide gel. The amount of DNA was quantified with a Qubit® 2.0 Fluorometer (Invitrogen, Carlsbad, CA, USA). The pooled library was diluted to a concentration of 26 pM. Template preparation was performed using the Ion PGM Template OT2 200 kit on Ion OneTouch™ 2 System (Thermo Fisher Scientific, Waltham, MA, USA) and the subsequent quality control was carried out on Qubit® 2.0 Fluorometer. The templates were sequenced on the Ion PGM™ System machine, using the Ion PGM sequencing 200 KIT V2 (Thermo Fisher Scientific, Waltham, MA, USA). QIIME 1.8.01 was used to process the sequence data [21].

Vaginal cytokine analysis

The analysis of a 48 cytokine and chemokine panel (including IL-1 β , IL-1ra, IL-2, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12(p70), IL-13, IL-15, IL-17, eotaxin, basic FGF, G-CSF, GM-CSF, IFN- γ , IP-10, MCP-1, MIP-1 α , PDGF-BB, MIP-1 β , RANTES, TNF- β , VEGF, IL-1 α , IL-2R α , IL-3, IL-12(p40), IL-16, IL-18, CTACK, GRO- α , HGF, IFN- α 2, LIF, MCP-3, M-CSF, MIF, MIG, β -NGF, SCF, SCGF- β , SDF-1 α , TNF- α , and TRAIL) was assessed in duplicate performed on vaginal swab using a magnetic bead-based multiplex immunoassays (Bio-Plex®) (BIO-RAD Laboratories, Milano, Italy) following manufacturers' instructions. In brief, 50 μ l of indiluted vaginal swab and standards was added in duplicate to a 96 multi-well plate containing analyte magnetic beads. After incubation for 30 min at room temperature, wells were then washed three times with Bio-Plex wash buffer, and 25 μ l of the premixed detection antibodies was added to the wells. This was followed by incubation for 30 min. After washing, 50 μ l of streptavidin-PE was added to the wells and the plate was incubated for 10 min with shaking. The

concentrations of the cytokines were determined using the Bio-Plex array reader (Luminex, Austin, TX). The data were analyzed using Bio-Plex Manager software (v.5, Bio-Rad) and were expressed as median fluorescence intensity (MFI) and concentration (pg/ml) [22].

Data availability The dataset was deposited in the Short Read Archive (PRJNA388787).

Results

A total of 20 patients with breast cancer were enrolled in the study. All of the patients had surgical therapy: 16 have subsequent hormonal therapy, 3 subsequent radiotherapy, and 1 chemo-radiotherapy.

The average age was 58.2 years, the average body mass index was 23.7 kg/m², and the average age of menarche was 12.4 years. Duration of menopausal status was 8.85 \pm 5.4 years.

Relevant demographic characteristics, pre-treatment clinical data, and inclusion/exclusion criteria were recorded at screening visit (T0). For each VRS parameter considered in the analysis for the VVA symptoms, a statistically significant improvement was recorded after the treatment except for dysuria (Table 1).

For Vaginal Health Index (VHI) and for the Female Sexual Function Index (FSFI), a statistically significant improvement was recorded after the laser treatment, while for the Female Sexual Distress Scale revised (FSDSr), no statistically significant improvement was recorded (Table 1).

We found higher levels of IL-18, CTACK, LIF, M-CSF, and IL-17 after treatment, while levels of IL-1ra, IL-2, IL-7, IL-9, IL-13, eotaxin, GM-CSF, and RANTES were lower after treatment (Table 2).

Levels of IL-1a, IL-2Ra, IL-3, IL-12p40, IL-16, GROa, HGF, IFN-a2, MCP-3, MIG, b-NGF, SCF, SCGF-b, SDF-1a, TNF-b, TRAIL, IL-1b, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12(p70), FGF-basic, G-CSF, IFN-g, IP-10, MCP-1(MCAF), MIP-1a, PDGF-bb, MIP-1b, and TNF-1 did not statistically change after laser treatment (Table 2).

Regarding the relative proportion of bacteria in the vaginal environment, the Shannon diversity index *H* and equitability comparison before and after treatment did not yield any statistical significant results (Table 3). The relative proportion of bacteria did not differ before and after treatment (Fig. 1).

Discussion

Micro-ablative fractional CO₂ method has been used as an effective tool for skin resurfacing and restoration [9], showing its remodeling connective tissue properties in many body

Table 1 Symptoms, signs, VHI, FSFI, and FSDSr before and after CO₂ laser treatment. *The variables are expressed as median (25th–75th percentile range)

| Symptoms | Before the fractional CO ₂ laser* | After the fractional CO ₂ laser* | <i>p</i> (non-parametric paired Wilcoxon test) |
|--------------------------|--|---|--|
| Vaginal dryness | 2 [2–3] | 2 [1–2] | 0.002 |
| Dyspareunia | 2 [2–3] | 2 [1–2] | 0.006 |
| Vaginal itching | 1 [0–2.75] | 0 [0–1] | 0.012 |
| Vaginal burning | 1 [0–3] | 1 [0–1] | 0.012 |
| Dysuria | 0 [0–1.75] | 0 [0–0.75] | 0.132 |
| Flattening vaginal folds | 2 [1.25–2.75] | 1 [1–1] | 0.000 |
| Vaginal mucosa dryness | 2 [2–2.75] | 1 [0–1.75] | 0.000 |
| Vaginal pallor | 2 [2–3] | 1 [1–1] | 0.000 |
| Fragility of the mucosa | 2 [1–2] | 1 [0–1] | 0.000 |
| Petechiae | 1 [0–1.75] | 0 [0–1] | 0.005 |
| VHI | 12 [11–13] | 16 [15.25–18.00] | 0.000 |
| FSFI | 27.5 [4–54.50] | 43 [20.25–70.50] | 0.003 |
| FSDSr | 21 [10–28] | 15 [8.00–24.00] | 0.074 |

Table 2 Expression of immune mediators in vaginal swab pre- and post-treatment (pg/ml)

| Vaginal fluid cytokines | Pre-treatment | Post-treatment | <i>p</i> value |
|---|-----------------------------------|-------------------------------|----------------|
| Significantly upregulated cytokines after treatment | | | |
| IL-18 | 582.6 [251.01–1574.0725] | 1729.35 [609.425–7017.5435] | 0.15 |
| CTACK | 8.4150 [6.4125–11.5325] | 11.2850 [6.412–18.31] | 0.044 |
| LIF | 21.77 [14.8175–29.355] | 30.4050 [17.1125–44.5825] | 0.025 |
| M-CSF | 21.4 [14.9875–38.19] | 42.93 [15.5050–66.8975] | 0.028 |
| IL-17 | 2.255 [1.29–7.6625] | 9.47 [3.17–13.22] | 0.028 |
| Significantly downregulated cytokines after treatment | | | |
| IL-1ra | 11,163.31 [8608.1525–16,511.5675] | 9111.96 [5137.73–11,322.9675] | 0.04 |
| IL-2 | 1.1250 [0.91–1.3975] | 0.94 [0.63–0.94] | 0.018 |
| IL-7 | 1.39 [0.77–2.0250] | 0.48 [0.4050–0.7350] | 0.000 |
| IL-9 | 6.445 [3.915–8.5475] | 2.92 [2.1825–6.88] | 0.014 |
| IL-13 | 1.245 [0.9775–1.73] | 0.72 [0.6275–1.0275] | 0.006 |
| Eotaxin | 16.3850 [10.85–40.9075] | 12.34 [8.21–20.4625] | 0.010 |
| GM-CSF | 67.4450 [60.99–74.765] | 62.2350 [51.8678–65.9625] | 0.01 |
| RANTES | 155.68 [62.5075–662.9350] | 81.9450 [9.6275–163.1850] | 0.004 |
| Cytokines with no significant change after treatment | | | |
| IL-1a | 10.3850 [1.4325–32.8250] | 11.08 [1.62–82.8725] | 0.1 |
| IL-2Ra | 68.9550 [45.9100–87.7225] | 72.85 [48.0975–110.89] | 0.279 |
| IL-3 | 140.5650 [121.7900–184.3450] | 169.39 [104.1525–282.6275] | 0.117 |
| IL-12p40 | 174.65 [107.9375–409.3875] | 169.43 [23.3575–371.3050] | 0.627 |
| IL-16 | 86.23 [51.0775–134.6225] | 69.425 [42.3675–148.8450] | 0.970 |
| GROa | 507.5350 [259.9125–1209.9350] | 582.6150 [194.2825–1180.66] | 0.794 |
| HGF | 84.47 [25.5125–242.7950] | 84.855 [36.34–143.455] | 0.411 |
| IFN-a2 | 46.08 [40.68–57.2850] | 49.17 [40.6625–69.1850] | 0.093 |
| MCP-3 | 43.9050 [30.1450–81.94] | 59.3650 [21.58–161.4975] | 0.212 |
| MIG | 258.42 [249.055–696.2675] | 399 [272.21–1448.2225] | 0.332 |
| b-NGF | 1.96 [1.21–2.95] | 3.5 [1.66–7.2975] | 0.22 |
| SCF | 23.81 [12.61–41.8] | 49.1950 [20.7625–68.6250] | 0.28 |
| SCGF-b | 222.12 [151.4575–298.54] | 409.58 [155.915–538.0675] | 0.40 |
| SDF-1a | 133.05 [90.8975–182.84] | 174.5850 [92.3075–256.8375] | 0.212 |
| TNF-b | 1.63 [0.795–2.3575] | 2.22 [0.87–3.89] | 0.296 |
| TRAIL | 35.1750 [14.2825–72.0525] | 32.96 [18.03–61.505] | 0.823 |
| IL-1b | 1.5350 [0.68–10.2325] | 4.64 [0.915–16.505] | 0.370 |
| IL-4 | 0.2150 [0.17–0.27] | 0.155 [1.225–0.3725] | 0.763 |
| IL-5 | 0.46 [0.2025–0.6375] | 0.2 [0.1550–0.39] | 0.000 |
| IL-6 | 1.125 [0.7950–1.76] | 0.49 [0.45–1.2] | 0.351 |
| IL-8 | 244.39 [63.7–909.185] | 195.1150 [41.5525–543.715] | 0.550 |
| IL-10 | 2.125 [1.71–3.6675] | 1.74 [1.2675–3.4425] | 0.370 |
| IL-12(p70) | 1.9450 [1.38–4.77] | 1.775 [0.9725–6.7325] | 0.247 |
| FGF-basic | 11.305 [8.35–13.0025] | 13.5 [7.675–16.8650] | 0.575 |
| G-CSF | 13.94 [9.9375–22.6375] | 7.7950 [4.2925–16.2450] | 0.263 |
| IFN-g | 5.19 [4.27–9.65] | 3.93 [3.2050–6.4975] | 0.198 |
| IP-10 | 855.92 [320.9575–2162.1525] | 840.7 [288.3050–2288.1175] | 0.494 |
| MCP-1(MCAF) | 2.46 [2.1225–3.3275] | 2.22 [1.8–3.12] | 0.334 |
| MIP-1a | 0.94 [0.84–1.2025] | 0.96 [0.845–1.1475] | 0.654 |
| PDGF-bb | 16.77 [11.6875–24.7650] | 10.075 [7.4875–13.7075] | 0.33 |
| MIP-1b | 27.8850 [6.4650–44.1450] | 22.65 [3.7175–107.58] | 0.575 |
| TNF-a | 3.5 [2.9250–5.2075] | 2.79 [2.3950–3.8675] | 0.277 |

Table 3 Comparison of bacterial diversity between cohorts. Bacterial diversity values are given as mean \pm standard deviation. Alpha diversity was compared between groups by means of a non-parametric *t* test using the compare_alpha_diversity.py script of QIIME. *p* values are shown in the last column

| Index | Before treatment | After treatment | <i>p</i> value |
|--------------|------------------|-----------------|----------------|
| Shannon | 1.6 0.9 | 1.8 \pm 1 | 0.702 |
| Equitability | 0.31 \pm 0.15 | 0.34 \pm 0.17 | 0.787 |

regions, and the ability to stimulate the production of collagen and elastic fibers. Indications of GSM symptom alleviation and improvement of sexual function satisfaction following the CO₂ laser therapy have been reported, bringing benefits especially in patients who cannot use estrogen therapy. Moreover, when laser treatment was compared with intravaginal estriol, the benefits of the CO₂ laser therapy were more pronounced and longer lasting compared with estriol [12, 13].

Previous data have suggested that specific laser irradiation parameters induced specific cellular activities, as cellular proliferation and a cell signaling cascade for the production and release of growth factors and cytokines, inducing the anti-inflammatory effect on the local tissue [23, 24]. Considering

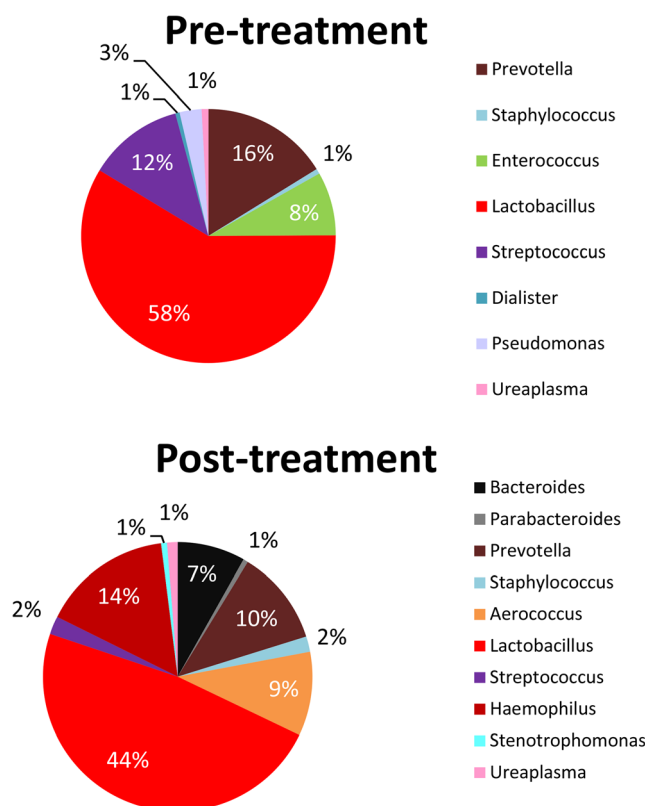


Fig. 1 The vaginal bacterial communities from patients before and after treatment. The output of plot_taxa_summary.py of QIIME showing the relative abundance of the predominant bacterial taxonomic groups in the studied cohorts

these findings, the aim of this study was to analyze the changes of vaginal cytokine profile and vaginal microbiome after CO₂ laser treatment and their possible role in improving symptoms of patients with VVA. A significant decrease in levels of pro-inflammatory cytokines, in particular IL-2 and IL-7, was observed after laser treatment, showing the action of this therapy on the inhibition of the release of certain inflammatory immune proteins. Another demonstration of the beneficial effect of this treatment is given by decrease of important inflammatory factors of cell-mediated immunity, such as RANTES, GM-CSF, and eotaxin, which attract monocytes, lymphocytes, basophils, and eosinophils in the inflammatory response. Conversely, the concentrations of some cytokines and growth factors, as IL-18, CTACK, LIF, and M-CSF, molecules usually involved in inflammatory processes, were found higher after laser therapy. Recent studies have shown that the laser irradiation on the skin increases cellular proliferation, viability of tissue, and remodeling of collagen. This mechanism induces an increase of inflammatory cytokine expression, pleiotropic proteins, which, in low concentrations, stimulate cell proliferation and migration, and they play an important role in tissue repair [29, 30]. Consistent with previous studies, the increase of concentration of IL-18, CTACK, LIF, and M-CSF proteins, in vaginal swab, could be the consequence of the remodeling status of the vaginal tissue after laser treatment. In particular, M-CSF is a growth factor involved in the pro-inflammatory immunologic mechanisms and it influences hematopoietic stem cells to differentiate into macrophages or other related cell types. M-CSF is particularly implicated in the steady-state control of tissue macrophage development and it has been proposed also to have a possible involvement in the resolution of inflammatory reactions. Following specific stimulations, this factor induces the differentiation of M2 macrophages which act as anti-inflammatory agents and resolve the inflammatory response not allowing more recruitment of other inflammatory cells and proceed in the tissue regeneration phase [31]. Prignano et al. [30] have conducted a study to verify the effects of fractional CO₂ laser on cytokine pathway of tissue repair of atrophic skin. The discrepancy observed in our study, where we did not find any statistically significant difference in b-FGF and PDGF expression after laser treatment, may be basically caused by two factors: (1) the different tissues involved (skin vs vaginal mucosa) and (2) the technique used for immune mediator analysis (immunohistochemical vs Bio-Plex platform) and laser parameters (different laser fluences).

It is well known that estrogen therapy is effective for the treatment of VVA, and, concomitantly, hormonal replacement increases the abundance of vaginal *Lactobacillus* spp. However, the independent contribution of the increase of *Lactobacilli* in improving VVA symptoms is not known. Data from Brotman provide evidence that there are differences in the bacterial communities between pre-, peri-, and

postmenopause, hypothesizing that the anaerobes may play a role in signs and symptomatology of VVA [32]. Instead, data from Shen [33] differ from those of Brotman et al. in so far as *Gardnerella* and *Atopobium* were the most frequent and abundant taxa in vaginal communities of women with AV and there was a statistically positive correlation between their prevalence and genital symptom scores. These compelling findings imply that strains of various bacterial species might be associated with the emergence of AV and that new therapeutic targets might emerge as more is learned about the possible causes of AV. In our data, the three bacterial genera *Lactobacillus*, *Prevotella*, and *Aerococcus* were within the five most represented in vaginal swab of pre- and post-treatment. *Streptococcus* and *Enterococcus* were more abundant before treatment while after treatment, *Haemophilus* and *Bacteroides* increased (Fig. 1). However, these differences of vaginal bacteria composition did not reach statistical significance.

Our study suggests that the beneficial effect of CO₂ laser treatment is primarily due to the biochemical and morphological changes of the epithelial vaginal cells which is associated with the expression of specific cytokines involved, basically, in anti-inflammatory process. Conversely, no change in composition of vaginal microbiome was documented. This may be related to the maintenance of a positive local balance able to favor the colonization of commensal microorganisms [16].

At our knowledge, this is the first study evaluating the effects of fractional CO₂ laser treatment in GSM by comparing the vaginal immunity with the vaginal microbiome in breast cancer women.

Clinical practice

In breast cancer survivors, atrophic vaginitis/vaginal dryness can affect up to 70% of the postmenopausal patients when compared to postmenopausal women without breast cancer, leading to the genitourinary syndrome of menopause (GSM).

After surgery, breast cancer survivors receive further chemo-, hormonal, and radiotherapy, which can aggravate the vaginal symptoms.

Recent reports suggest that laser therapy may be valuable as a non-hormonal therapeutic modality in the management of GSM.

Preliminary results of laser therapy efficacy have been reported on specific symptoms of GSM, such as sexual function and vulvovaginal atrophy, vaginal flora, and GSM as a whole.

We also observed a significant reduction of clinical symptoms related to GSM after the CO₂ laser treatment, supporting its efficacy as therapy in postmenopausal cancer survivors. The high remodeling status in vaginal epithelium is demonstrated by the significant changes in inflammatory and modulatory cytokine patterns. The absence of modifications in the

vaginal microbiome after the treatment confirms the safety of the CO₂ laser. The availability of randomized controlled trials and well-designed case-control studies are required to further investigate the potential benefits, harm, and efficacy of laser therapy in the treatment of GSM symptoms.

Conclusion

The significant reduction of clinical symptoms related to GSM after the CO₂ laser treatment supports its efficacy as therapy in postmenopausal cancer survivors. The high remodeling status in vaginal epithelium is demonstrated by the significant changes in inflammatory and modulatory cytokine patterns. The absence of modifications in the vaginal microbiome after the treatment confirms the safety of the CO₂ laser. The exact mechanism responsible for the clinical improvement is still to be elucidated and further studies are needed in this field to clarify its long-term efficacy and other effects. The availability of randomized controlled trials and well-designed case-control studies are required to further investigate the potential benefits, harm, and efficacy of laser therapy in the treatment of GSM symptoms.

Authorship Conceptualization, M.C.; methodology, A.M.B., G.C., and N.Z.; software, G.C. and N.Z.; investigation, A.M.B., G.C., and N.Z.; resources, Z.T., S.G., F.P., A.P., and G.S.; data curation, G.C. and N.Z.; writing—original draft, G.C., N.Z., and M.C.; writing—review and editing, M.C.; visualization, G.C. and N.Z.; funding acquisition, M.C.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no competing interests.

Ethics approval and consent to participate The study was approved by the Ethics Committee of Florence. The study was conducted in accordance with the Declaration of Helsinki.

Consent for publication All patients who were recruited for the study signed an informed consent form.

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